***In vitro* transcription**

T7 promoter containing transcription templates were generated by PCR using Go Tag enzyme (Promega): Fas exon 6 WT/noBP templates were generated from Fas WT minigene, Fas M0 template was generated from Fas M0 minigene (Izquierdo et al, 2007), Fas exon 6 TAG mutant template was generated from a ssDNA oligonucleotide (see table X for all sequences). PCR products were purified on agarose gel.

Cy5-CTP/Cy5-UTP labeled RNA were transcribed directly from the PCR templates using Megascript T7 Transcription kit (Ambion) accordingly to the manufacturer’s instructions.

**A complex formation**

15 ng/ul fluorescently labeled RNA were incubated with 3 ul of HeLa cell nuclear extracts (CILBIOTECH) supplemented with 3 mM MgCl2, 24.9 mM KCl, 3.33% PVA, 13.3 mM HEPES pH 8, 0.13 mM EDTA,13.3 % glycerol, 0.03 % NP-40, 0.66 mM DTT, 2 mM ATP and 22 mM creatine phosphate in a final volume of 9 ul. The mixture was incubated for 18 min at 30 oC. 1 ul of heparin (10 ug/ul stock) was added and incubated for 10 min at room temperature. 3 ul of 50% glycerol were added and 9 ul loaded on a composite gel (4% acrylamide, 0.05% bis-acrylamide, 0.5% agarose, 50mM Tris, 50mM glycine). The gel was run for 6 hours at 200 Volts in 50mM Tris / 50mM glycine buffer. After electrophoresis, fluorescence was detected using a Typhoon PhosphorImager.

**U1 snRNP / U2 snRNP inactivation**

The inactivation of U1 snRNP and U2 snRNP was performed as described in Donmez et al, Mol Cell, 2007 using 2′-O-methylated oligoribonucleotide complementary to U1 snRNA (5’-CUGCCAGGUAAGUAU-3’) or U2 snRNA (5’-CAGAUACUACACUUG-3’).

t-test, tails =2, type 2

from 20 replicates from 7 experiments for RNA wt and from 19 replicates from 6 experiments for RNA TAG.